

Effect of Household Cooking Methods on Polyphenols and Antioxidant Capacity of Endogenous Leafy Vegetables

Carole Nadia Adjouavi SOSSA-VIHOTOGBE¹, Houédougbe Noël AKISSOE², Bienvenu Victor ANIHOUVI², Djidjoho Joseph HOUNHOUIGAN²

¹Faculté d'Agronomie, Université de Parakou, Parakou, République du Bénin

²Ecole de Nutrition des Sciences et Technologies Alimentaires, Faculté des Sciences Agronomiques, Université d'Abomey-Calavi, O3BP2819 Jericho, Cotonou, République du Bénin.



Abstract – Leafy vegetables are often suggested to pretreatments prior to use. One of the usual practice is the cooking methods which presumably affect biochemical and physicochemical compounds. This study aims to evaluate various cooking methods of leafy vegetables, namely precooking without additive, cooking with additive (traditional potash called Kanmu or ash filtrate) or cooking without additive on total polyphenols content and antioxidant activity of three traditional leafy vegetables (*Justicia tenella*, *Acmella uliginosa* and *Sesamum radiatum*). Total polyphenol contents (TPC) ranged between 0.36 – 0.60 g GAE/100g DW. Traditional cooking methods affected biochemical and physicochemical compounds depending on species. For instance, precooking and cooking with potash decreased significantly ($p < 0.05$) TPC in *J. tenella* leaves while no significant effect was evidenced in the case of cooking with ash filtrate and cooking without additive. Regarding antioxidant capacity, precooking and cooking without additive significantly increased it [raw 39.01 – 63.9%), P and CWA (85.58 – 90.89%)] whilst cooking with additive decreased it (CP, CF; 10.77-44.44%).

Keywords – *Justicia tenella*, *Acmella uliginosa*, *Sesamum radiatum*, bioactives compounds.

I. INTRODUCTION

Leafy vegetables are usually consumed after some culinary treatments like squeeze washing, pre-cooking or cooking [1], [2] and these treatments provoke contrasting physico-chemical changes, depending on the species. [3] reported losses in antioxidant components, particularly low antioxidant activity of broccoli (*Brassica oleracea*, Brassicaceae) while other studies mentioned that cooking increased antioxidant components of leafy vegetables [4], [5].

In Benin, processing techniques of traditional leafy vegetables before consumption were documented [2]. Irrespective of culinary treatments [6], [1], [2], some additives such as ash filtrate and alkaline rock named 'kanmu' or 'kanwa' was usually used [2]. Details regarding the nutritional impacts, essentially biochemical and physicochemical changes of these practices are not fully documented. Some interactions between nutritional constituents and additive probably impact bioavailability of nutrients. As already pointed out, the cooking of *Vernonia amygdalina* leaves with kanwa decreased significantly phenolic compounds, saponin and oxalic acid [1]. Due to the mineral potential of additives used (ash filtrate, 'kanmu'), pre-treatments associating these additives can presumably impact nutritional and bioactive constituents of leafy vegetables.

This study aims to evaluate the effect of household cooking methods on total phenolic compounds and antioxidant capacity of leafy vegetables: *Sesamum radiatum* L. (Black benniseed, Pedaliaceae), *Acmella uliginosa* (L.) Jansen (Toothache plant, Asteraceae) and *Justicia tenella* (Nees) T. Anderson (Acanthaceae).

II. MATERIAL AND METHODS

2.1- Sample collection

Samples of traditional leafy vegetables were collected in markets, farms and traditional garden of villages in the Department of Donga in North-West Benin (in Natitingou, Pam-pam, Perma, Tiélé). After collection, edible part was immediately separated from branches, put in sealed polyethylene bag, and in cooler with ice before returning in laboratory. Alkaline rock (kanmu or kanwa) was bought from Natitingou market and ash filtrate was obtained by filtration of sorghum stem ash collected in three households.

2.2- Processing treatments and materials preparation for analysis

For each species, sample of 100 g of leaf was cleaned with distilled water before being submitted to different treatments. The quantities of water and softening ingredients used were indicated by housewives in the sampling areas. Water temperature before leaves addition was set at $90^{\circ}\text{C} \pm 3^{\circ}\text{C}$. Processing methods of all leafy vegetables were summarized on figures 1 and 2 based on household techniques. Spinaches (*A. uliginosa* and *J. tenella*) were cooked following three scenarios: (i) cooking with potash (alkali rock: CP) or ash filtrate (CF); (ii) cooking without additive (CWA) and (iii) pre-cooking without additive (P). Regarding *S. radiatum*, only scenarios 1 and 2 (CP/CF, CWA) were applied.

After cooking, samples and raw leaves were freeze-dried (-55°C), then milled in Moulinex (Waring, 32B780) and stored in airtight containers for analysis.

2.3- Physicochemical analyses

Total phenolic content

Total phenolic content (TPC) was determined following the modified method of [7] by [8] and [9] to determine TPC in leafy vegetables flour fruitfully. Extraction was realized twice by centrifugation using HCl/methanol (1%v/v). Gallic acid (Aldrich Chemical Company Steinheim, Germany) was used as standard and the optical density was measured at 760 nm with a spectrophotometer (Jenway, UV 6715). Data was expressed in Gallic Acid Equivalent (GAE)/ 100g Dry Weight (DW).

Antioxidant capacity

DPPH (2,2-diphenyl-1-picrylhydrazyl acid) method was used to assess antioxidant capacity of raw and cooked leafy vegetables following the modified method of [10] used by [3]. 25 mg of freeze-dried flour of leafy was extracted with 1.125 ml of aqueous methanol (80%) during 2 h under continuous stirring at room temperature twice. The supernatant was filtrated with Whatman N°1 paper and diluted with 5% aqueous methanol.

Statistical analysis

One-way Analysis of Variance was performed using STATISTICA (v6) program to evaluate effect of cooking methods on total

phenolics and antioxidant capacity. The effect of traditional cooking methods was compared to raw leafy using the test of Newman and Keuls at 5% to structure mean values.

III. RESULTS

3.1- Knowledge on the processing of studied leafy vegetable

A result of survey to know cooking parameters for studied leafy vegetables was indicated in table 1. It was noticed that, cooking parameters were different according to species and household's methods.

Table 1: Cooking conditions of fresh leaves using different cooking methods (100 g)

Species	Cooking modes	Cooking time (min)	Quantity of water (ml)	Quantity of potash/ash filtrate (g)
<i>Sesamum radiatum</i>	CP/CF	9	915	5.9/26.7
	CWA	9	915	-
<i>Justicia tenella</i>	CP/CF	9	610	5.50/7.3
	CWA	9	610	-
	P	3	1200	-
<i>Acmella uliginosa</i>	CP/CF	5	713	3.3/23.7
	CWA	5	713	-
	P	7	1170	-

CP= Cooking with potash, CF= cooking with ash filtrate, CWA= cooking without additive, P= precooking

3.2- Total Polyphenolic content (TPC)

TPC of uncooked leafy was presented in Table 2. Precooking (P) and cooking with potash (CP) significantly decreased the TPC ($p < 0.05$) for *J. tenella*, while no significant effect was observed for cooking without additive (CWA) and cooking with ash filtrate (CF). In contrast, significant increase of TPC was observed after precooking for *A. uliginosa* but no significant effect was observed for other cooking methods (CP, CWA, and CF) for this species. As far as *S. radiatum* is concerned, no significant effect for all cooking method was observed on TPC.

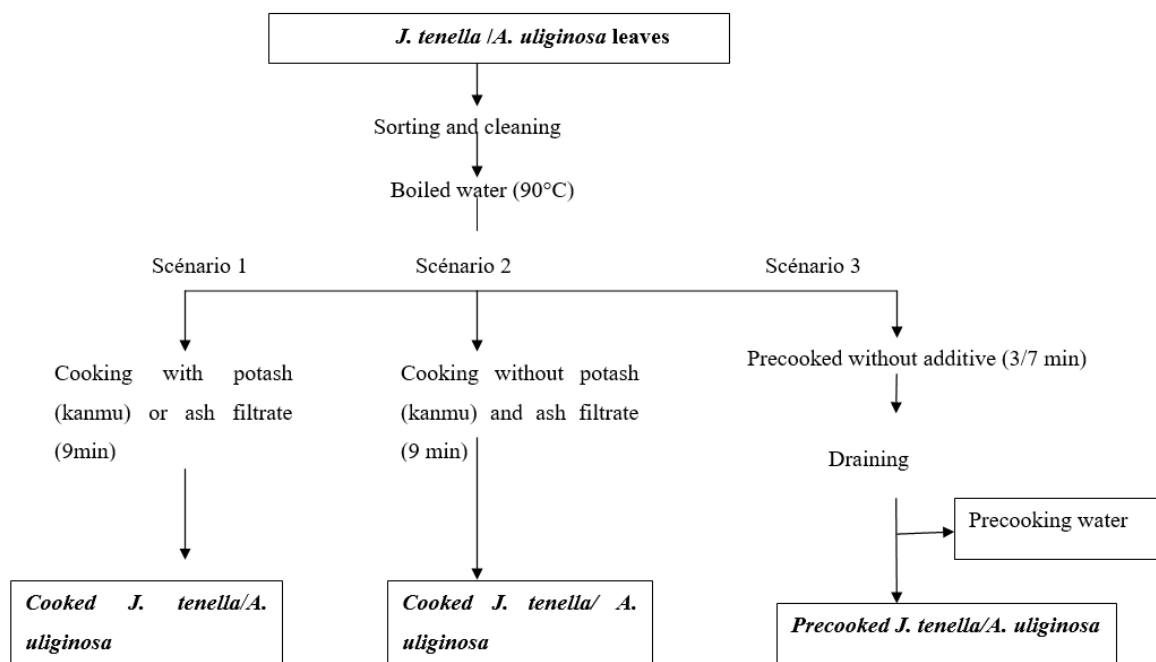


Figure 1: Processing methods applied for *Justicia tenella* and *Acmella uliginosa*.

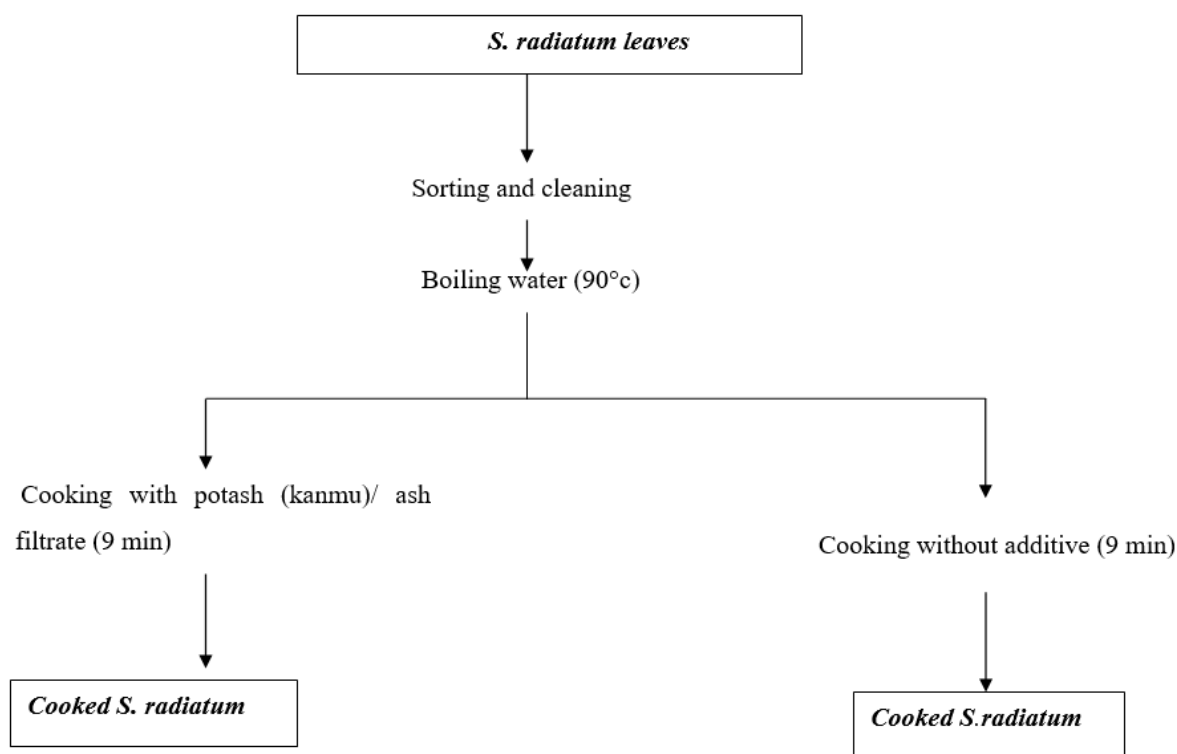


Figure 2: Processing methods applied for *Sesamum radiatum*

Table 2: Total phenolic compounds in leafy vegetables after different treatments

Parameters	Species	R	P	CWA	CP	CF
TPC (g GAE/100g DW)	<i>Justicia tenella</i>	0.56±0.12a	0.38±0.01b	0.59±0.12a	0.44±0.07bc	0.53±0.12ac
	<i>Acmella uliginosa</i>	0.36±0.05a	0.62±0.11b	0.43 ±0.04a	0.37±0.07a	0.43±0.08a
	<i>Sesamum radiatum</i>	0.60±0.15a	-	0.58±0.03a	0.53±0.20a	0.55±0.11a

Means ±SD, values with different letters on the same line are significantly different ($p < 0.05$), GAE: Gallic Acid Equivalent, R= raw, P= precooking, CWA= cooking without additive, CP= cooking with potash, CF= cooking with ash filtrate

Table 3: Antioxidant capacity of leafy vegetables during cooking

Antioxidant capacity	Species	C	P	CSA	CP	CF
DPPH (%)	<i>Justicia tenella</i>	74.08±7.62b	63.22±18.1ab	52.95±16.63a	3.78±2.08c	6.25±22.32d
	<i>Acmella uliginosa</i>	39.01±6.51c	85.58±4b	88.26±2.09b	15.84±14.16a	22.16±8.52a
	<i>Sesamum radiatum</i>	63.9±34 a	-	90.89±0.3c	0.77±7.16b	44.44±25.6a

Results of ANOVA + Newman and Keuls test: Means ±SD, values with different letters on the same line that have different letters are significantly different ($p < 0.05$), R= raw, P= precooking, CWA= cooking without additive, CP= cooking with potash, CF= cooking with ash filtrate

3.3- Effect of cooking methods on antioxidant capacity

Inhibition percent of DPPH radical varied between 3.78 - 74.08%, 15.84 - 39.01% and 10.77 - 90.89% for *J. tenella*, *A. uliginosa* and *S. radiatum*, respectively (Table 4). A significant decrease of antioxidant activity was recorded during treatments for *J. tenella* ($p < 0.05$). Only precooking significantly increased antioxidant activity of *A. uliginosa*, while only cooking without additive significantly increased it in the case of *A. uliginosa* and *S. radiatum*.

IV. DISCUSSION

The high variability of TPC was associated to leafy species and it showed two antagonist behaviors depending on the treatments. An increase of TPC in *A. uliginosa* (72.2%) and *J. tenella* (5.35%) during precooking was reported during cooking of *Spinacia oleracea*, *Petroselinum Crispum*, *Anethum graveolens* and other traditional leafy vegetables such as *Ocimum gratissimum*, *Talinum triangulare*, *Amaranthus hybridus*, *Brassica juncea*, *Brassica rapa* and *Solanum retroflexum* Dum [4], [5], [11], [12], [13]. This increase might be rooted in the breakdown of cell wall and liberation of TPC that is more quantified [14]. The raise of free flavonols content subsequent to cooking could also explain this increase [15]. Conversely, a significant decrease of TPC was noticed during heat treatments of fruits, leafy vegetables, sorghum and legumes [16], [17], [18], [19], [4], [20], [8], [21], [22]. A decrease induced by precooking method could be explained in one hand by leakage of some TPC or number of flavonoids affected by heat treatment in boiling water [23], [24] or by breakdown of molecule link during cooking [25]. In other hand, TPC decrease could be due to the formation of hydroxyl groups during cooking and formation of insoluble complex with minerals or proteins; in this form, identification of reactive hydroxyl groups could decrease [26].

In general, tenderizer or additives induce a downward trend for TPC, probably due to their linkage with polyphenol's hydroxyl groups [27]. Actually, traditional tenderizer had variable mineral composition such as Ca, Fe, Zn and Na [27]. The Kanmu used in this study contains 94, 82 and 0.054 mg/ 100g DW of calcium, iron and zinc, respectively.

The DPPH percent inhibition ranged between 39.0 and 74.1% and lower than that of BHA (93.6%), one synthetic antioxidant. Culinary effect varied within species. Antioxidant activity decreased in the case of *J. tenella* or increased after precooking of *A. uliginosa* or cooking with tenderizer. Wachtel-Galor *et al.* [28] in the case of Brassicaceae also mentioned this variability of antioxidant activity as a result of cooking methods. Antioxidant activity decreased from 14.65 to 94.95% and, this is in line with [3] and [11] who reported the decrease of antioxidant activity after cooking during 2-3 and 6 min. Amin *et al.* [29] also mentioned that blanching reduced *Amaranthus* leaves antioxidant activity. The reduction of scavenging activity after cooking with potash or ash filtrate could be affected by samples pH. Actually, the pH of leaves obtained with ash filtrate ranged between 9.02 and 9.91 while those obtained with potash varied from 10.15 to 10.30. [30] found that antioxidant activity decreased significantly when the pH is 10 and it is low at pH=10 than pH= 9. As far as the increase in scavenging activity (during *A. uliginosa* precooking, *A. uliginosa* and *S. radiatum* cooking without additive) is concerned, we are agreement with other research works [31]. Turkmen *et al.* [4] obtained the same trend after cooking of pepper and green beans. Boiling of plantain, pumpkin, mushrooms and leafy vegetables [32], [5], [33], [34] increased their antioxidant activity. According to [35], this rise could be due to breakage of the cell wall and cellular compartments that release more antioxidant components or due to chemical reactions with production of antioxidants compounds. The increase of antioxidant activity in cooked leaves could be also due to inactivation of oxidative enzymes by heating [36]. According to [37], enhance of scavenging activity after tomato cooking could be caused by increase of discharge of phyto-chemical substances while [31] attributed the increase of activity in cooked leaves to formation of novel products with antioxidant activity.

V. CONCLUSION

Beside a large variation nutritional composition of studied species, different cooking processes had contrasting physicochemical changes depending on species. Actually, precooking affects significantly TPC content of some of leafy vegetables. Cooking with potash increase significantly TPC content of *J. tenella* and nitrate in all studied leafy vegetables., ash filtrate does not influence significantly TPC, in all species. In contrast, cooking with additive decrease significantly antioxidant capacity of all studied species. Antioxidant activity of all species is reduced during cooking with tenderizer. The effect of traditional cooking methods on vitamins content could be explored to understand their contribution in reduction antioxidant capacity. It will be important to identify the appropriate cooking methods that decrease antinutritional content and preserve vitamins and mineral.

ACKNOWLEDGEMENT

We thank International Foundation of Science (IFS) for financial support for this research.

REFERENCES

- [1] Ejoh R.A, Nkonga U.V., Gouado I., Mbofung C.M. (2007). Nutritional Components of Some Non-Conventional Leafy Vegetables Consumed in Cameroon. *Pakistan Journal of Nutrition*, 6(6):712-717.
- [2] Sossa-Vihotogbé C.N.A., Akissoe H.N., Anihouvi V.B., Ahohuendo B.C. Ahanchede A., Sanni A., Hounhouigan D.J. (2012). Endogenous knowledge of four leafy vegetables used by rural populations in Benin. *Ecology of Food and Nutrition*, 51(1):22-39.
- [3] Zhang D., Hamauzu Y. (2004). Phenolics, ascorbic acid, carotenoids and antioxidant activity of broccoli and their changes during conventional and microwave cooking. *Food Chemistry*, 88:503-509.
- [4] Turkmen N., Sari F., Velioglu S.Y. (2005). The effect of cooking methods on total phenolics and antioxidant activity of selected green vegetables. *Food Chemistry*, 93:713-718.
- [5] Adefegba S.A., Oboh G. (2011). Cooking enhances the antioxidant properties of some tropical green leafy vegetables. *African Journal of Biotechnology*, 10(4):632-639.
- [6] Dansi A., Adjatin A., Adoukonou-Sagbadja H., Faladé V., Yedomonhan H., Odou D., Dossou B. (2008). Traditional leafy vegetables and their use in the Benin Republic. *Genetic Resources and Crop Evolution*, 55(8):1239-1256.
- [7] Singleton V.L., Rosii J.A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*, 16:144-158.
- [8] Kayodé P., Linnemann A.R.Jr., Nout M., AJS van Boekel M. (2007). Impact of sorghum processing on phytate, phenolic

- compounds and in vitro solubility of iron and zinc in thick porridges. *Journal of the Science of Food and Agriculture*, 87:832-838.
- [9] Sossa-Vihotogbe C.N.A., Akissoé N.H., Anihouvi B.V., Ahohuendo B.C., Ahanchede A. Hounhouigan J.D (2013). Effect of fertilization and harvesting time on antioxidant activity of three leafy vegetables commonly used in Benin, *African Journal of Food, Agriculture, Nutrition and Development*. 13(5): 1-20.
- [10] Brand-Williams W., Cuvelier M.E., Berset C. (1995). Use of a free radical method to evaluate antioxidant activity. *Lebensmittel. Wissenschaft und Technologie*, 28:25-30.
- [11] Kamel S.M. (2013) Effect of microwave treatments on some bioactive compounds of parsley (*Petroselinum Crispum*) and dill (*Anethum graveolens*) leaves. *Journal of Food Processing and Technology*, 4:233. doi:10.4172/2157-7110.1000233
- [12] Khwairakpan B., Balwinder S., Kiran B. (2014). Effect of domestic cooking methods on the nutritive and antioxidative components of mustard leaves (*Brassica juncea*). *International Journal of Food, Agriculture and Veterinary Sciences*, 4 (1):24-31.
- [13] Managa M.G., Shai J., P A.D.T., Sultanbawa Y., Sivakuma D. (2020). Impact of household cooking techniques on African Nightshade and Chinese Cabbage on phenolic compounds, antinutrients, in vitro antioxidant and β -glucosidase activity. *Frontiers in nutrition*, 1-13.
- [14] Oboh G., Rocha J.B.T. (2007). Polyphenols in red pepper [*Capsicum annum* var *aviculare* (Tepin)] and their protective effect on some pro-oxidant induced lipid peroxidation in brain and liver. *European Food Research and Technology*, 225:239-247.
- [15] Stewart A.J., Bozonnet S., Mullen W., Jenkins G.I., Michael E.J., Crozier A. (2000). Occurrence of flavonols in tomatoes and tomato-based products. *Journal of Agricultural and Food Chemistry*, 48:2663-2669.
- [16] Towo E.E., Svanberg U.L.F., Ndossi G.D. (2003). Effect of grain pretreatment on different extractable phenolics groups in cereals and legumes commonly consumed in Tanzania. *Journal of the science of Food and Agriculture*, 83:98-986.
- [17] Ismail A., Marjan Z.M., Foong C.W. (2004). Total antioxidant activity and phenolic content in selected vegetables, *Food Chemistry*, 87:581-586.
- [18] Sahlin E., Savage G.P., Lister C.E. (2004). Investigation of the antioxidant properties of tomatoes after processing. *Journal of Food Composition and Analysis*, 17:635-647.
- [19] Zhang D., Hamazu Y. (2004). Phenolics, ascorbic acid, carotenoids and antioxidant activity of broccoli and their changes during conventional and microwave cooking. *Food Chemistry*, 88:503-509.
- [20] Finocchiaro F., Ferrari B., Gianinetti A., Dall'Asta C., Galaverna G., Seazzina F., Pellegrini N. (2007). Characterization of antioxidant compounds of red and white rice and changes in total antioxidant capacity during processing. *Molecular Nutrition and Food Research*, 51(8): 1006-1019.
- [21] Liu W., Zu Y., Fu Y., Kong Y., Mac W., Yang M., Li, J., Wua N. (2010). Variation in contents of phenolic compounds during growth and postharvest storage of pigeon pea seedlings. *Food Chemistry*, 121:732-739.
- [22] Oulai P.D., Zoue L.T., Bedikou M.E., Megnanou R-M., Niamke S.L. (2014). Impact of cooking on nutritive and antioxidant characteristics of leafy vegetables consumed in northern Côte d'Ivoire. *International Journal of Plant, Animal and Environmental Sciences*, 4(3):576-585.
- [23] Trichopoulou A., Vasilopoulou E., Hollman P., Chamalides C., Foufa E., Kaloudis T., Kromhout D., Miskaki P., Petrochilou I., Poulima E., Stafilaskis K., Theophilou D. (2000). Nutritional composition and flavonoid content of edible wild greens and green pies: a potential rich source of antioxidant nutrients in the Mediterranean diet. *Food chemistry*, 70:319-323.
- [24] Yadav S.K., Sehgal S. (2003). Effect of domestic processing and cooking on selected antinutrient contents of some green leafy vegetables. *Plant Foods for Human Nutrition*, 58:1-11.
- [25] Crozier A., Lean M.E.J., McDonald M.S., Black C. (1997). Quantitative analysis of the flavonoid content of commercial

- tomatoes, onions, lettuce, and celery. *Journal of Agricultural and Food Chemistry*, 45:590-595.
- [26] Matuschek E., Towo E., Svanberg U. (2001). Oxidation of polyphenols in phytate reduced high-tannin cereals: Effect on different phenolic groups and on in vitro available iron. *Journal of Agricultural Food Chemistry*, 49:5630–5638.
- [27] Eyzaguirre R.Z., Nienaltowska K., de Jong L. E. Q., Hasenack B. B. E., Nout M. J. R. (2006). Effect of food processing of pearl millet (*Pennisetum glaucum*) IKMP.5 on the level of phenolics, phytate, iron and zinc. *Journal of Science Food and Agriculture*, 86:1391-1398.
- [28] Wachtel-Galor S., Wong K.W., Benzie I.F.F. (2008). The effect of cooking on Brassica vegetables. *Food Chemistry*, 110:706-710.
- [29] Amin I., Norazaidah Y., Hainida K.I.E. (2006). Antioxidant activity and phenolic content of raw and blanched *Amaranthus* species. *Food Chemistry*, 94:47-52.
- [30] Settharaksa S., Jongjareonrak A., Hmadhlu P., Chansuwan W., Siripongvutikorn S. (2012). Flavonoid, phenolic contents and antioxidant properties of Thai hot curry paste extract and its ingredients as affected of pH, solvent types and high temperature. *International Food Research journal*, 19:1581-1587.
- [31] Nicoli M.C., Anese M., Parpinel M. (1999). Influence of processing on the antioxidant properties of fruit and vegetables. *Trends in Food Science and Technology*, 10: 94-100.
- [32] Azizah A.H., Wee K.C., Azizah O., Azizah M. (2009). Effect of boiling and stir frying on total phenolics, carotenoids and radical scavenging activity of pumpkin (*Cucurbita moschato*). *International Food Research Journal*, 16:45-51.
- [33] Ebun O., Santosh K. (2011). Effect of domestic cooking on the polyphenolic content and antioxidant capacity of plantain (*Musa paradisiaca*). *World Journal of Dairy and Food Sciences*, 6(2):189-194.
- [34] Sun L., Zhuang Y., Bai X. (2011). Effects of boiling and microwaving treatments on nutritional characteristics and antioxidant activities of *Agaricus blazei* Murril. *International Journal of Food Science and Technology*, 46:1209-1215.
- [35] Maeda H., Katsuki T., Akaike T., Yasutake R. (1992). High correlation between lipid peroxide radical and tumor promoter effect: Suppression of tumor promotion in the Epstein-Barr virus/B-lymphocyte system and scavenging of alkyl peroxide radicals by various vegetable extracts. *Japanese Journal of Cancer Research*, 83:923-928.
- [36] Yamaguchi T., Mizobuchi T., Kajikawa R., Kawashima H., Miyabe F., Terao J., Takamura H., Matoba T. (2001). Radical Scavenging activity of vegetables and the effect of cooking on their activity. *Food Science and Technology Research*, 7(3):250-257.
- [37] Gahler S., Otto K., Bohm V. (2003). Alterations of vitamin C, total phenolics and antioxidant capacity as affected by processing tomatoes to different products. *Journal of Agricultural and Food Chemistry*, 51, 7962-7968.